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Terms	Documents
L3 and (HCV envelope protein)	9

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<u>L4</u>	L3 and (HCV envelope protein)	9	<u>L4</u>
<u>L3</u>	L2 and vaccinia	9769	<u>L3</u>
<u>L2</u>	435/320.1.ICLS.	32624	<u>L2</u>
<u>L1</u>	(435/320.1ICLS.)![IPC]	0	<u>L1</u>

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NEWS	19	FEB 26	MEDLINE reloaded with enhancements
NEWS	20	FEB 26	EMBASE enhanced with Clinical Trial Number field
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NEWS	23	FEB 26	CAS Registry Number crossover limit increased from 10,000 to 300,000 in multiple databases
NEWS	24	MAR 15	WPIDS/WPIX enhanced with new FRAGHITSTR display format
NEWS	25	MAR 16	CASREACT coverage extended
NEWS	26	MAR 20	MARPAT now updated daily
NEWS	27	MAR 22	LWPI reloaded
NEWS	28	MAR 30	RDISCLOSURE reloaded with enhancements
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NEWS	30	APR 02	JICST-EPLUS removed from database clusters and STN
NEWS EXPRESS			NOVEMBER 10 CURRENT WINDOWS VERSION IS V8.01c, CURRENT MACINTOSH VERSION IS V6.0c(ENG) AND V6.0Jc(JP), AND CURRENT DISCOVER FILE IS DATED 25 SEPTEMBER 2006.
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=> vaccinia

L1 21398 VACCINIA

=> HCV adj envelope

L2 0 HCV ADJ EVELOPE

=> "HCV envelope protein"

L3 184 "HCV ENVELOPE PROTEIN"

=> L1 and L3

L4 6 L1 AND L3

=> D L4 IBIB ABS 1-6

L4 ANSWER 1 OF 6 CAPLUS COPYRIGHT 2007 ACS on STN

ACCESSION NUMBER: 2001:912918 CAPLUS

DOCUMENT NUMBER: 137:150837

TITLE: Effect of downstream sequence on the cleavage of envelope protein 1 signal sequence in Hepatitis C virus

AUTHOR(S): Zhu, Lixin; Kong, Yuying; Wang, Yuan; Li, Guangdi

CORPORATE SOURCE: Institute of Biochemistry and Cell Biology, Shanghai Institute for Biological Sciences, Chinese Academy of Sciences, Shanghai, 200031, Peop. Rep. China

SOURCE: Shengwu Huaxue Yu Shengwu Wuli Xuebao (2001), 33(6), 682-686

CODEN: SHWPAU; ISSN: 0582-9879

PUBLISHER: Shanghai Kexue Jishu Chubanshe

DOCUMENT TYPE: Journal

LANGUAGE: Chinese

AB The RNA genome of hepatitis C virus encodes a polyprotein of about 3,000 amino acids, which is processed into 10 viral protein by proteases provided by host cells and virus itself, multiple precursors are produced due to inefficient processing. E1 signal sequence (C/E1 site) processing in eukaryotic vaccinia virus/T7 system was studied. Differently truncated HCV structural proteins were expressed in this system. It was found that the efficient cleavage of E1 signal sequence was affected by downstream envelope protein sequences. When the lacZ gene encoding a

product with similar size was engineered downstream to the E1 signal sequence, the inefficient cleavage of signal sequence was also observed, suggesting that the effect of downstream sequence on the cleavage was due to the presence of the envelope protein sequences. Computer-aided analysis clearly showed that E1 signal sequences was a typical signal sequence. The influence of downstream sequences to signal sequence cleavage demonstrated here was uncommon. To date, similar observations were only reported for the processing of IL-12 signal sequence and the C/prM site of flavivirus.

L4 ANSWER 2 OF 6 CAPLUS COPYRIGHT 2007 ACS on STN

ACCESSION NUMBER: 1999:117461 CAPLUS
DOCUMENT NUMBER: 130:324135
TITLE: New monoclonal antibodies against a recombinant second envelope protein of hepatitis C virus.
AUTHOR(S): Inudoh, Michiharu; Kato, Nobuyuki; Tanaka, Yuetsu
CORPORATE SOURCE: Virology Division, National Cancer Center Research Institute, Chuo-ku, Tokyo, 104-0045, Japan
SOURCE: Microbiology and Immunology (1998), 42(12), 875-877
CODEN: MIIMDV; ISSN: 0385-5600
PUBLISHER: Center for Academic Publications Japan
DOCUMENT TYPE: Journal
LANGUAGE: English

AB To study the immunol. features of the hepatitis C virus (HCV) envelope protein (E2 protein), new specific monoclonal antibodies (mAbs) were generated. WKA/H rats were immunized with syngeneic cells infected with a vaccinia virus expressing the E2 protein and with soluble E2 protein obtained from Chinese hamster ovary cells with a plasmid-based expression system. By screening hybridoma cells obtained from spleen cells of the immunized rats, three specific mAbs were obtained. One mAb was reactive to a peptide corresponding to the hypervariable region 1 (HVR1) in E2 protein, while the others reacted to regions outside HVR1. The significance of these antibodies for the diagnosis of HCV infection as well as for anal. of the structure of the HCV E2 protein will be discussed.

REFERENCE COUNT: 10 THERE ARE 10 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L4 ANSWER 3 OF 6 CAPLUS COPYRIGHT 2007 ACS on STN

ACCESSION NUMBER: 1994:4188 CAPLUS
DOCUMENT NUMBER: 120:4188
TITLE: Characterization of hepatitis C virus envelope glycoprotein complexes expressed by recombinant vaccinia viruses
AUTHOR(S): Ralston, Robert; Thudium, Kent; Berger, Kim; Kuo, Carol; Gervase, Barbara; Hall, John; Selby, Mark; Kuo, George; Houghton, Michael; Choo, Qui Lim
CORPORATE SOURCE: Chiron Corp., Emeryville, CA, 94608, USA
SOURCE: Journal of Virology (1993), 67(11), 6753-61
CODEN: JOVIAM; ISSN: 0022-538X
DOCUMENT TYPE: Journal
LANGUAGE: English

AB The authors constructed recombinant vaccinia virus vectors for expression of the structural region of hepatitis C virus (HCV). Infection of mammalian cells with a vector (vv/HCV1-906) encoding C-E1-E2-NS2 generated major protein species of 22 kDa (C), 33 to 35 kDa (E1), and 70 to 72 kDa (E2), as observed previously with other mammalian expression systems. The bulk of the E1 and E2 expressed by vv/HCV1-906 was integrated into endoplasmic reticulum membranes as core-glycosylated species, suggesting that these E1 and E2 species represent intracellular forms of the HCV envelope proteins. HCV E1 and E2 formed E1-E2 complexes which were precipitated by either anti-E1 or

anti-E2 serum and which sedimented at approx. 15 S on glycerol d. gradients. No evidence of intermol. disulfide bonding between E1 and E2 was detected. E1 and E2 were copurified to approx. 90% purity by mild detergent extraction, followed by chromatog. on Galanthus nivalus lectin-agarose and DEAE-Fractogel. Immunization of chimpanzees with purified E1-E2 generated high titers of anti-E1 and anti-E2 antibodies. Further studies demonstrated that purified E1-E2 complexes were recognized at high frequency by HCV+ human sera and generated protective immunity in chimpanzees, suggesting that these purified HCV envelope proteins display native HCV epitopes.

L4 ANSWER 4 OF 6 CAPLUS COPYRIGHT 2007 ACS on STN

ACCESSION NUMBER: 1992:528131 CAPLUS

DOCUMENT NUMBER: 117:128131

TITLE: Hepatitis C virus asialoglycoproteins manufacture for vaccines or immunoassay

INVENTOR(S): Ralston, Robert O.; Marcus, Frank; Thudium, Kent B.; Gervase, Barbara A.; Hall, John A.

PATENT ASSIGNEE(S): Chiron Corp., USA

SOURCE: PCT Int. Appl., 28 pp.

CODEN: PIXXD2

DOCUMENT TYPE: Patent

LANGUAGE: English

FAMILY ACC. NUM. COUNT: 8

PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
WO 9208734	A1	19920529	WO 1991-US8272	19911107
W: AU, CA, CS, FI, HU, JP, NO, PL, RO, SU				
RW: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LU, NL, SE				
EP 414475	A1	19910227	EP 1990-309120	19900821
EP 414475	B1	19971210		
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AT 161041	T	19971215	AT 1990-309120	19900821
ES 2110411	T3	19980216	ES 1990-309120	19900821
CA 2064705	A1	19910226	CA 1990-2064705	19900822
CA 2064705	C	19990406		
WO 9102820	A1	19910307	WO 1990-US4766	19900822
W: AU, CA, JP				
AU 9063449	A	19910403	AU 1990-63449	19900822
AU 655156	B2	19941208		
JP 05502156	T	19930422	JP 1990-512531	19900822
JP 2001314192	A	20011113	JP 2001-75114	19900822
WO 9115771	A1	19911017	WO 1991-US2225	19910329
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RW: BF, BJ, CF, CG, CM, GA, ML, MR, SN, TD, TG				
AU 9176510	A	19911030	AU 1991-76510	19910329
AU 639560	B2	19930729		
GB 2257784	A	19930120	GB 1992-20480	19910329
BR 9106309	A	19930420	BR 1991-6309	19910329
HU 62706	A2	19930528	HU 1992-3146	19910329
HU 217025	B	19991129		
JP 05508219	T	19931118	JP 1991-507636	19910329
JP 2733138	B2	19980330		
RO 109916	B1	19950728	RO 1975-92012	19910329
PL 172133	B1	19970829	PL 1991-296329	19910329
RU 2130969	C1	19990527	RU 1991-5053084	19910329
EP 450931	A1	19911009	EP 1991-302910	19910403
EP 450931	B1	19960612		
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EP 693687	A1	19960124	EP 1995-114016	19910403
EP 693687	B1	19990728		
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AT 139343	T	19960615	AT 1991-302910	19910403
ES 2088465	T3	19960816	ES 1991-302910	19910403
AT 182684	T	19990815	AT 1995-114016	19910403
ES 2134388	T3	19991001	ES 1995-114016	19910403
CA 2095521	A1	19920509	CA 1991-2095521	19911107
CA 2203443	A1	19920509	CA 1991-2203443	19911107
CA 2203443	C	20010828		
AU 9190267	A	19920611	AU 1991-90267	19911107
AU 668078	B2	19960426		
EP 556292	A1	19930825	EP 1992-900091	19911107
EP 556292	B1	19991229		
EP 556292	B2	20061129		
R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE				
JP 06504431	T	19940526	JP 1992-500944	19911107
HU 66063	A2	19940928	HU 1993-1336	19911107
EP 842947	A2	19980520	EP 1997-120661	19911107
EP 842947	A3	20011212		
EP 842947	B1	20040421		
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RU 2123528	C1	19981220	RU 1993-43621	19911107
PL 175610	B1	19990129	PL 1991-300038	19911107
AT 188220	T	20000115	AT 1992-900091	19911107
ES 2139591	T3	20000216	ES 1992-900091	19911107
RO 115446	B1	20000228	RO 1993-626	19911107
JP 2001286290	A	20011016	JP 2001-59335	19911107
CZ 289006	B6	20011017	CZ 1993-824	19911107
RU 2175657	C2	20011110	RU 1997-115378	19911107
JP 2003093081	A	20030402	JP 2002-199317	19911107
JP 2003174875	A	20030624	JP 2002-353148	19911107
EP 1471073	A2	20041027	EP 2004-76119	19911107
EP 1471073	A3	20041201		
R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE				
FI 106317	B1	20010115	FI 1992-4349	19920928
NO 9203839	A	19921119	NO 1992-3839	19921001
NO 310241	B1	20010611		
FI 107803	B1	20011015	FI 1993-2025	19930505
NO 9301680	A	19930628	NO 1993-1680	19930507
NO 304380	B1	19981207		
LV 10344	B	19960220	LV 1993-4381	19930531
US 5679342	A	19971021	US 1993-97853	19930727
LT 3808	B	19960325	LT 1993-1747	19931230
US 5968775	A	19991019	US 1995-438435	19950510
US 5712087	A	19980127	US 1995-440519	19950512
US 6312889	B1	20011106	US 1995-440549	19950512
FI 9701702	A	19970421	FI 1997-1702	19970421
FI 107804	B1	20011015		
NO 9702213	A	19970514	NO 1997-2213	19970514
NO 304381	B1	19981207		
PT 102022	B	20001229	PT 1997-102022	19970626
CZ 289923	B6	20020417	CZ 1997-2196	19970710
JP 11071395	A	19990316	JP 1998-103178	19980414
JP 3207155	B2	20010910		
GR 3031361	T3	20000131	GR 1999-402455	19990929
GR 3032771	T3	20000630	GR 2000-400473	20000228
JP 2004049235	A	20040219	JP 2003-180211	20030624
JP 2005187479	A	20050714	JP 2005-35317	20050210
JP 2006219503	A	20060824	JP 2006-145982	20060525
PRIORITY APPLN. INFO.:			US 1989-398667	A 19890825
			US 1990-611419	A 19901108

US 1990-611965	A 19901108
US 1991-758880	A 19910913
US 1987-122714	B2 19871118
US 1987-139886	B2 19871230
US 1988-161072	B2 19880226
US 1988-191263	B2 19880506
US 1988-263584	B2 19881026
US 1988-271450	B2 19881114
US 1989-325338	B2 19890317
US 1989-341334	B2 19890420
US 1989-353896	B2 19890421
US 1989-355002	B2 19890518
US 1989-355961	B2 19890518
US 1989-456637	B2 19891221
US 1990-504352	A 19900404
JP 1990-512531	A3 19900822
JP 2001-75114	A3 19900822
WO 1990-US4766	A 19900822
WO 1991-US2225	A 19910329
EP 1991-302910	A3 19910403
CA 1991-2095521	A3 19911107
CZ 1993-824	A3 19911107
EP 1992-900091	A3 19911107
EP 1997-120661	A3 19911107
JP 1992-500944	A3 19911107
JP 1998-103178	A3 19911107
JP 2001-59335	A3 19911107
JP 2002-199317	A3 19911107
WO 1991-US8272	A 19911107
US 1992-910760	A3 19920707
FI 1993-2025	A 19930505
US 1993-97853	A1 19930727
JP 2005-35317	A3 20050210

AB Two hepatitis C virus (HCV) envelope proteins (E1 and E2) are manufactured without sialylation. Expression of these genes in lower eukaryotes, or in mammalian cells in which terminal glycosylation is blocked, results in proteins similar to native HCV glycoproteins. When isolated by mannose-binding GNA (Galanthus nivalus agglutinin) lectin affinity, the E1 and E2 proteins aggregate into virus-like particles. Cells bearing a mannose receptor or asialoglycoprotein receptor are capable of being infected with HCV and of supporting culturing of the virus. E1 and E2 were produced in HeLa S3 cells inoculated with recombinant Vaccinia virus containing HCV gene fragments and purified using a GNA-agarose column.

L4 ANSWER 5 OF 6 BIOSIS COPYRIGHT (c) 2007 The Thomson Corporation on STN
 ACCESSION NUMBER: 1999:49108 BIOSIS
 DOCUMENT NUMBER: PREV199900049108
 TITLE: New monoclonal antibodies against a recombinant second envelope protein of hepatitis C virus.
 AUTHOR(S): Inudoh, Michiharu; Kato, Nobuyuki; Tanaka, Yuetsu [Reprint author]
 CORPORATE SOURCE: Dep. Biosci., Sch. Sci., Kitasato Univ., Kitasato 1-15-1, Sagami-hara, Kanagawa 228-8555, Japan
 SOURCE: Microbiology and Immunology, (1998) Vol. 42, No. 12, pp. 875-877. print.
 CODEN: MIIMDV. ISSN: 0385-5600.
 DOCUMENT TYPE: Article
 LANGUAGE: English
 ENTRY DATE: Entered STN: 10 Feb 1999
 Last Updated on STN: 10 Feb 1999

AB To study the immunological features of the hepatitis C virus (HCV

) envelope protein (E2 protein), new specific monoclonal antibodies (mAbs) were generated. WKA/H rats were immunized with syngeneic cells infected with a vaccinia virus expressing the E2 protein and with soluble E2 protein obtained from Chinese hamster ovary cells with a plasmid-based expression system. By screening hybridoma cells obtained from spleen cells of the immunized rats, three specific mAbs were obtained. One mAb was reactive to a peptide corresponding to the hypervariable region 1 (HVR1) in E2 protein, while the others reacted to regions outside HVR1. The significance of these antibodies for the diagnosis of HCV infection as well as for analysis of the structure of the HCV E2 protein will be discussed.

L4 ANSWER 6 OF 6 BIOSIS COPYRIGHT (c) 2007 The Thomson Corporation on STN
ACCESSION NUMBER: 1993:585942 BIOSIS
DOCUMENT NUMBER: PREV199497005312
TITLE: Characterization of hepatitis C virus envelope glycoprotein complexes expressed by recombinant vaccinia viruses.
AUTHOR(S): Ralston, Robert; Thudium, Kent; Berger, Kim; Kuo, Carol; Gervase, Barbara; Hall, John; Selby, Mark; Kuo, George; Houghton, Michael [Reprint author]; Choo, Qui-Lim
CORPORATE SOURCE: Chiron Corporation, 4560 Horton St., Emeryville, CA 94608, USA
SOURCE: Journal of Virology, (1993) Vol. 67, No. 11, pp. 6753-6761. CODEN: JOVIAM. ISSN: 0022-538X.
DOCUMENT TYPE: Article
LANGUAGE: English
ENTRY DATE: Entered STN: 28 Dec 1993
Last Updated on STN: 28 Dec 1993

AB We constructed recombinant vaccinia virus vectors for expression of the structural region of hepatitis C virus (HCV). Infection of mammalian cells with a vector (vv/HCV-1-906) encoding C-E1-E2-NS2 generated major protein species of 22 kDa (C), 33 to 35 kDa (E1), and 70 to 72 kDa (E2), as observed previously with other mammalian expression systems. The bulk of the E1 and E2 expressed by vv/HCV-1-906 was found integrated into endoplasmic reticulum membranes as core-glycosylated species, suggesting that these E1 and E2 species represent intracellular forms of the HCV envelope proteins. HCV E1 and E2 formed E1-E2 complexes which were precipitated by either anti-E1 or anti-E2 serum and which sedimented at approximately 15 S on glycerol density gradients. No evidence of intermolecular disulfide bonding between E1 and E2 was detected. E1 and E2 were copurified to approximately 90% purity by mild detergent extraction followed by chromatography on Galanthus nivalus lectin-agarose and DEAE-Fractogel. Immunization of chimpanzees with purified E1-E2 generated high titers of anti-E1 and anti-E2 antibodies. Further studies, to be reported separately, demonstrated that purified E1-E2 complexes were recognized at high frequency by HCV+ human sera (D. Y. Chien, Q.-L. Choo, R. Ralston, R. Spaete, M. Tong, M. Houghton, and G. Kuo, Lancet, in press) and generated protective immunity in chimpanzees, -(Q.-L. Choo, G. Kuo, R. Ralston, A. Weiner, D. Chien, G. Van Nest, J. Han, K. Berger, K. Thudium, J. Kansopon, J. McFarland, A. Tabrizi, K. Ching, B. Mass, L. B. Cummins, E. Muchmore, and M. Houghton, submitted for publication), suggesting that these purified HCV envelope proteins display native HCV epitopes.